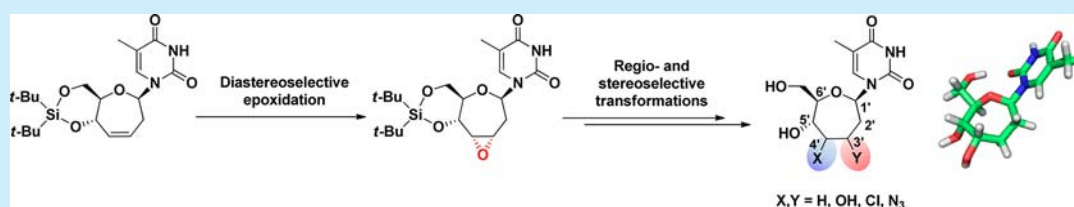


Seven-Membered Ring Nucleoside Analogues: Stereoselective Synthesis and Studies on Their Conformational Properties

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Supporting Information



ABSTRACT: The synthesis of a novel series of seven-membered ring nucleoside analogues as candidates for biological screening and gene silencing applications is described. The key step in the synthetic approach is a stereoselective synthesis of an epoxide that is used as a common synthetic intermediate to prepare functionalized oxepane nucleoside derivatives. The conformational landscape and preferred ring-puckering of selected oxepane nucleosides was also studied by NMR, X-ray crystallography, and quantum mechanical calculations.

Nucleoside analogues are extremely valuable compounds in medicinal chemistry.^{1,2} Modification of the furanose moiety and its impact on the conformation and biological activity of nucleosides has been studied extensively.^{3,4} Due to their more challenging accessibility, ring-expanded derivatives have been considerably less explored. Hexose- and hexitol-based nucleosides are the most studied among this class and were first described by the Eschenmoser^{5,6} and Herdewijn^{7,8} groups, respectively. In the hexitol series (1), the flexible furanose ring is replaced with a more rigid six-membered sugar ring, and the position of the nucleobase is moved from the anomeric carbon to the 3'-position (Figure 1). Nucleic acids derived from hexitol nucleosides have been proven useful for gene silencing, biotechnology, and synthetic biology applications.⁹

Seven-membered oxepane scaffolds are commonly found in natural products and biologically active molecules.^{10–13} In the search for ring-expanded DNA analogues, we recently reported the synthesis of oligonucleotides constructed from oxepane nucleosides (oxepane nucleic acids, ONA, 2).¹⁴ It was hypothesized that expanding the carbohydrate moiety of DNA to a seven-membered skeleton would provide conformationally more flexible sugars relative to the six-membered ring pyranose, thus better mimicking the conformation of natural 2-deoxyribose. Indeed, ONA, like DNA oligonucleotides, are able to trigger RNase H-mediated degradation of a complementary RNA strand.¹⁴ Following our work, a few more

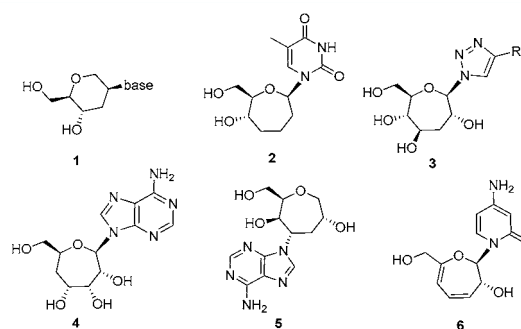


Figure 1. Hexitol nucleosides (1), oxepane nucleoside previously synthesized in our laboratory (2), other examples of oxepane nucleosides reported to date (3–6).

examples of oxepane nucleoside analogues have been reported as potential glycosidase inhibitors (3),¹⁵ antiviral agents (4¹⁶ and 5¹⁷), and unexpected byproducts of nucleoside syntheses (6)¹⁸ (Figure 1).

As part of our ongoing interest in the development of nucleosides with expanded sugar rings,^{14,19} we herein describe the synthesis of several novel oxepane-based nucleosides that are excellent candidates for biological screening and building blocks

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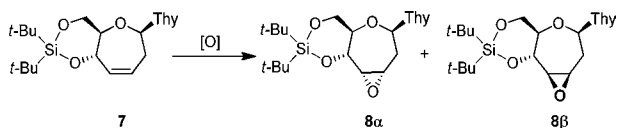
for gene silencing oligonucleotides.²⁰ The focus of this study is to use a common and versatile chiral synthetic intermediate to generate structurally diverse oxepane nucleosides.

We also report for the first time on the conformation of some of the synthesized analogues, assessed by X-ray crystallography, NMR, and theoretical (classical and quantum mechanical) calculations.

Synthesis and Characterization of Oxepane Nucleosides

An efficient five-step synthesis of oxepene **7** was previously described by our laboratory,¹⁴ starting from commercially available tri-*O*-acetyl-D-glucal and adapting the ring expansion strategy described by Hoberg et al.²¹ The attractiveness of **7** is that its alkene functionality can be manipulated for the introduction of different functional groups in the seven-membered ring. For example, access to a new series of functionalized oxepane nucleoside monomers was devised from an asymmetric epoxidation of **7**, enabling the diastereoselective installation of new stereocenters. Thus, we first focused on finding the best conditions for the stereoselective epoxidation of **7** (Scheme 1).

Scheme 1. Stereoselective Synthesis of Oxepane Nucleoside **8**



We attempted to take advantage of the allylic alcohol to perform an asymmetric epoxidation. Thus, the silyl bridge of **7** was cleaved before subjecting it to epoxidation with VO(acac)₂.²² Under the experimental conditions, we observed rapid decomposition of the starting material. Classical oxidation of **7** with *m*CPBA in DCM afforded a 2:1 α/β diastereomeric mixture of the epoxide in 50% yield. The two isomers showed very similar polarity (TLC, $\Delta R_f < 0.1$), making it difficult to separate them by silica gel column chromatography (Table 1, entry 1). Next, we

Table 1. Epoxidation of Protected Oxepene **7 under Different Conditions**

entry	epoxidation reagent	cat. (mol %)	yield (%)	$8\alpha/8\beta^f$	de (%)
1 ^a	<i>m</i> CPBA		50 ^d	2:1	33
2 ^b	(<i>S,S</i>)-Jacobsen cat.	5	36 ^d	1:1.7	26
3 ^b	(<i>R,R</i>)-Jacobsen cat.	5	34 ^e	14:1	87
4 ^c	(<i>R,R</i>)-Jacobsen cat.	10	58 ^e	14:1	87

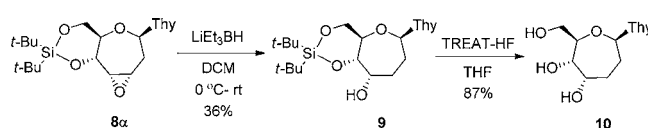
Reaction conditions: ^aTHF, 40 °C; ^bNaOCl, DCM, 4-phenylpyridine *N*-oxide (^b20 mol %, ^c40 mol %), rt. ^dCrude epoxide yield ($\alpha + \beta$). ^eIsolated yields for **8 α** . ^fRatio calculated by NMR in the isolated crude mixture.

used a chiral catalyst to induce stereoselectivity. Jacobsen catalyst is an attractive candidate due to its ability to catalyze stereoselective epoxidations of unfunctionalized olefins. When **7** was treated with 5 mol % of (*S,S*)-Jacobsen catalyst in a NaOCl/CH₂Cl₂ biphasic system and in the presence of pyridine *N*-oxide, a 1:1.7 α/β diastereomeric mixture was obtained (Table 1, entry 2). A significant improvement in stereoselectivity (87% de) was observed when the same reaction was carried out with the (*R,R*)-Jacobsen catalyst under the same conditions (Table 1, entry 3). The reaction proceeded in better yields (58%) by increasing the catalyst loading to 10 mol % without any decrease in stereoselectivity (Table 1, entry 4). Traces of **8 β** were removed

by flash column chromatography. It is noteworthy that, at this stage, the configuration of the epoxide ring was ambiguous as the NOESY NMR spectra did not show the expected key correlations. However, subsequent formation of the anhydro oxepane **12** from **8 α** (Scheme 3) allowed the assignment of stereochemistry at C3'–C4'.

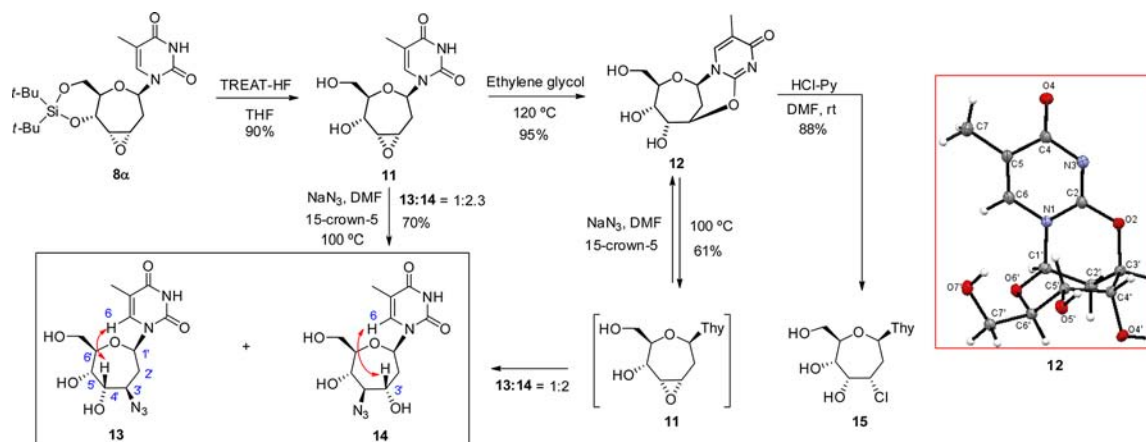
The oxepane scaffold was then ready for diversification via epoxide opening to generate functionalized oxepane nucleosides. The reaction of **8 α** with 10 equiv of LiEt₃BH at 0 °C resulted in a completely stereo- and regioselective conversion to **9** in 36% yield, together with unreacted starting material. Additional equivalents of LiEt₃BH did not increase the yield of the reaction and resulted in formation of base reduction products. Regioselective opening of epoxide **8 α** can be explained by the possible coordination of LiEt₃BH to the nucleobase and directing the attack of hydride to the proximal and less hindered C3' position. The regio- and stereochemistry of **9** was confirmed by COSY and NOESY NMR experiments, respectively. Cleavage of the silyl protecting group afforded nucleoside **10** in 87% yield (Scheme 2).

Scheme 2. Regio-/Stereoselective Synthesis of Oxepane Nucleoside **10**



To further explore the scope of our strategy to synthesize structurally diverse oxepane nucleosides, diastereomerically pure **8 α** was desilylated with TREAT-HF in 90% yield to afford epoxide **11** that was then reacted with NaN₃ in DMF at 100 °C and in the presence of 15-crown-5 (Scheme 3). Unlike the hydride reduction described above for protected nucleoside **8 α** , the azidation reaction of **11** resulted in a 1:2.3 mixture of **13** and **14** in 70% combined yield. The regioselectivity and yield of this step dropped significantly when azidation was carried out on **8 α** instead. The position of the azide group in both **13** and **14** regioisomers was determined by COSY NMR in DMSO-*d*₆. The configuration was assessed by NOESY experiments. A clear correlation between H6 of the nucleobase and H3' was observed for compound **14**, suggesting the β configuration of H3'. In the case of regioisomer **13**, a correlation between H4' and H6 was observed instead, suggesting a *cis* relative orientation between the thymine and H4' (Scheme 3).

Interestingly, reaction of epoxide **11** with NaF/KHF₂ in ethylene glycol at 120 °C resulted in an isomerization to afford anhydro nucleoside **12** in high yield, as assessed by NMR and UV experiments. NaF/KHF₂ salts were found to not play a role in the isomerization, as heating **11** in the absence of fluoride afforded **12** in the same yield and reaction time. The identity, stereochemistry, and conformation of anhydro nucleoside **12** were unambiguously assigned by single-crystal X-ray diffraction. Within the bicyclic structure, the oxepane ring adopts a chairlike conformation in which the hydroxymethyl group and 5'-OH are equatorial, while the thymine base and the 4'-OH group are pseudoaxial (Scheme 3). We next turned our attention to chlorine functionalization, given the recent discovery of chlorinated nucleoside analogues with promising biological activity.^{23,24} Indeed, reaction of **12** with HCl-Py in DMF provided the desired chlorinated nucleoside analogue **15** with complete stereoselectivity in 88% yield (Scheme 3).

Scheme 3. Synthesis of Azido (13, 14) and Chloro (15) Oxepane Nucleosides^a

^aCrystal structure of nucleoside 12 and key NOE correlations in 13 and 14.

Interestingly, when 12 was subjected to azidation conditions, the expected product resulting from nucleophilic α -attack at the 3'-position was not detected. Instead, a regioisomeric mixture of compounds 13 and 14 was obtained in a ratio similar to that of azidation of epoxide 11. As previously reported for furanose nucleosides,²⁵ we hypothesize that 12 equilibrates to epoxide 11 under these conditions, and that 11 is the direct precursor of 13 and 14 (Scheme 3). To further study the mechanism of this reaction, 12 was dissolved in DMF-*d*₆ in the absence of NaN₃ and crown ether. After being heated for 3 h at 100 °C in an NMR tube, no changes were observed in the ¹H NMR spectrum, suggesting that nucleophilic conditions are crucial in this case to drive the equilibrium toward epoxide 11.

Conformational Analysis

With this new set of oxepane nucleosides in hand, we next embarked on the analysis of the structural properties of two of the analogues, 10 and 11. The ring pucker of seven-membered ring systems can be described as a combination of boat/twist-boat (B/TB) and chair/twist-chair (C/TC) pseudorotational cycles.^{26,27} The torsional landscape of 10 and 11 was first assessed by molecular dynamics simulations coupled to a metadynamics enhanced sampling scheme (see Supporting Information). The most stable conformations were further minimized using quantum mechanical density functional theory calculations at the M062X/6-31+G(d,p) level using MeOH as solvent (full details in Supporting Information). Our calculations show that lowest-energy states for epoxide 11 correspond to a chairlike conformation, in which all the substituents assume a pseudoequatorial disposition to alleviate steric interactions (Figure 2a). The differences in stability among the lowest-energy states are due to small variations in the orientation of the thymine and hydroxymethyl substituents; meanwhile, the sugar pucker remains almost invariable (details in Supporting Information). Lowest-energy states for the alcohol derivative 10 are mixtures of twisted chair and boat conformations (~65 and ~35%, respectively) (Figure 2b). The main differences among low-energy states are also in this case due to small variations in the orientation of the substituents, causing very minor variations in the sugar conformation (Supporting Information).

Calculated ³J_{HH} values based on the Boltzmann distribution of the computed low-energy conformers of 10 and 11 were compared with experimental ³J_{HH} values from ¹H NMR coupling

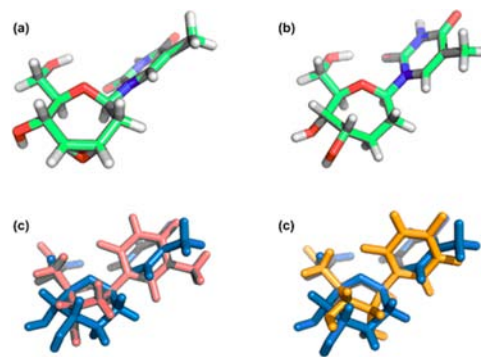


Figure 2. Lowest-energy conformations of (a) 11 and (b) 10. Derivative 10 is a mixture of boat and twisted boat conformations, and 11 is found exclusively in the chair conformation. Superposition of 10 (blue) with (c) C2'-endo-thymidine (red) and (d) C3'-endo-uridine (orange).

constant analysis in MeOH-*d*₄ (Table 2; details in Supporting Information). For 11, there was sufficient spectral resolution of all the resonances so that coupling constants could be conveniently extracted from the spectra. In the case of 10, the large number of couplings involved in H2' and H3' protons prevented the extraction of the coupling constants between these

Table 2. Calculated and Experimental ³J_{H,H} Coupling Constants (±0.2 Hz) for 10 and 11

coupling constant	calcd 10 ^a	exptl 10 ^b	calcd 11 ^a	exptl 11 ^b
³ J _{1',2'}	11.0	10.0	10.1	10.5
³ J _{1',2''}	2.3	3.4	0.7	2.4
³ J _{2',3'}	1.7		0.3	0
³ J _{2',3''}	12.3		5.7	4.5
³ J _{2',3'}	6.0		NA	NA
³ J _{2',3''}	2.5		NA	NA
³ J _{3',4'}	1.0	2.0	NA	NA
³ J _{3',4''}	9.9	9.8	4.0	4.7
³ J _{4',5'}	4.2	2.5	0.6	0
³ J _{5',6'}	7.9	5.7	9.1	9.6

^aValues calculated using GIAO/B3LYP/aug-cc-pVDZ methodology on optimized structures, averaged based on their Boltzmann population. ^bObserved coupling constants from ¹H NMR experiments in MeOH-*d*₄.

protons. In general, there was good correspondence between calculated and observed $^3J_{\text{HH}}$, supporting the accuracy of the conformations provided by computation. The H1' splitting pattern in both **10** and **11** are consistent with the pseudoequatorial orientation of the pyrimidine base. Figure 2c,d compares the 3D structure of compound **10** to the natural nucleosides (dT and rU) by superposition of the structures. When the bases are aligned, the pseudoequatorial C4'–OH bond in **10** superimposes rather well with the pseudoequatorial C3'–OH bond of uridine (shown in the favored C3'-endo conformation), suggesting that 4',7'-linked oligomers of **10** may conform to the structure of natural nucleic acids (e.g., RNA).

In summary, we have described the synthesis of several novel oxepane nucleosides by using a diastereomerically pure epoxide as a common intermediate. Theoretical calculations and NMR studies suggest that these analogues adopt a chair conformation with the pyrimidine base in the equatorial orientation. We expect this new series of ring-expanded nucleic acid analogues to be useful as building blocks in the design of new functional genetic systems and gene silencing oligonucleotides.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.orglett.5b02769.

General methods, experimental procedures, full spectroscopic data, summary of energies, geometries, and details of classical and quantum mechanical calculations for each conformer of nucleosides **10** and **11** (PDF)
Crystallographic information for **12** (CIF)

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Notes

The authors declare no competing financial interest.

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■ REFERENCES

- (1) Sofia, M. J. *Annu. Rep. Med. Chem.* **2014**, *49*, 221–247.
- (2) Parker, W. B. *Chem. Rev.* **2009**, *109*, 2880–2893.
- (3) Marquez, V. E.; Ben-Kasus, T.; Barchi, J. J.; Green, K. M.; Nicklaus, M. C.; Agbaria, R. *J. Am. Chem. Soc.* **2004**, *126*, 543–549.
- (4) Marquez, V. E.; Ezzitouni, A.; Russ, P.; Siddiqui, M. A.; Ford, H.; Feldman, R. J.; Mitsuya, H.; George, C.; Barchi, J. J. *J. Am. Chem. Soc.* **1998**, *120*, 2780–2789.
- (5) Pitsch, S.; Wendeborn, S.; Jaun, B.; Eschenmoser, A. *Helv. Chim. Acta* **1993**, *76*, 2161–2183.
- (6) Eschenmoser, A.; Dobler, M. *Helv. Chim. Acta* **1992**, *75*, 218–259.

(7) Verheggen, I.; Van Aerschot, A.; Toppet, S.; Snoeck, R.; Janssen, G.; Balzarini, J.; De Clercq, E.; Herdewijn, P. *J. Med. Chem.* **1993**, *36*, 2033–2040.

(8) Aerschot Van, A.; Verheggen, I.; Hendrix, C.; Herdewijn, P. *Angew. Chem., Int. Ed. Engl.* **1995**, *34*, 1338–1339.

(9) Taylor, A. L.; Pinheiro, V. B.; Smola, M. J.; Morgunov, A. S.; Peak-Chew, S.; Cozens, C.; Weeks, K. M.; Herdewijn, P.; Holliger, P. *Nature* **2015**, *518*, 427–430.

(10) Kanojia, R. M.; Wachter, M. P.; Levine, S. D.; Adams, R. E.; Chen, R.; Chin, E.; Cotter, M. L.; Hirsch, A. F.; Huettemann, R. *J. Org. Chem.* **1982**, *47*, 1310–1319.

(11) Basu, S.; Ellinger, B.; Rizzo, S.; Deraeve, C.; Schürmann, M.; Preut, H.; Arndt, H.-D.; Waldmann, H. *Proc. Natl. Acad. Sci. U. S. A.* **2011**, *108*, 6805–6810.

(12) Berger, D.; Overman, L. E.; Renhowe, P. A. *J. Am. Chem. Soc.* **1993**, *115*, 9305–9306.

(13) Basu, S.; Waldmann, H. *Bioorg. Med. Chem.* **2014**, *22*, 4430–4444.

(14) Sabatino, D.; Damha, M. J. *J. Am. Chem. Soc.* **2007**, *129*, 8259–8270.

(15) Castro, S.; Cherney, E. C.; Snyder, N. L.; Peczu, M. W. *Carbohydr. Res.* **2007**, *342*, 1366–1372.

(16) Sizun, G.; Dukhan, D.; Griffon, J.-F.; Griffe, L.; Meillon, J.-C.; Leroy, F.; Storer, R.; Sommadossi, J.-P.; Gosselin, G. *Carbohydr. Res.* **2009**, *344*, 448–453.

(17) Tripathi, S.; Roy, B. G.; Drew, M. G. B.; Achari, B.; Mandal, S. B. *J. Org. Chem.* **2007**, *72*, 7427–7430.

(18) Nomura, M.; Endo, K.; Shuto, S.; Matsuda, A. *Tetrahedron* **1999**, *55*, 14847–14854.

(19) Abou Assi, H.; Martínez-Montero, S.; Dixit, D. M.; Chua, Z.; Bohle, D. S.; Damha, M. J. *Eur. J. Org. Chem.* **2015**, *2015*, 1945–1953.

(20) Deleavey, G. F.; Damha, M. J. *Chem. Biol.* **2012**, *19*, 937–954.

(21) Hoberg, J. O. *J. Org. Chem.* **1997**, *62*, 6615–6618.

(22) Itoh, T.; Jitsukawa, K.; Kaneda, K.; Teranishi, S. *J. Am. Chem. Soc.* **1979**, *101*, 159–169.

(23) Srivastav, N. C.; Shakya, N.; Mak, M.; Liang, C.; Tyrell, D. L. J.; Agrawal, B.; Kumar, R. *Bioorg. Med. Chem.* **2010**, *18*, 7542–7547.

(24) Srivastav, N. C.; Shakya, N.; Bhavanam, S.; Agrawal, A.; Tse, C.; Desroches, N.; Kunimoto, D. Y.; Kumar, R. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 1091–1094.

(25) Al-Masoudi, N. A.; Pfeleiderer, W. *Carbohydr. Res.* **1995**, *275*, 95–105.

(26) Jahn, M. K.; Dewald, D. A.; Vallejo-López, M.; Cocinero, E. J.; Lesarri, A.; Zou, W.; Cremer, D.; Grabow, J.-U. *Chem. - Eur. J.* **2014**, *20*, 14084–14089.

(27) DeMatteo, M. P.; Snyder, N. L.; Morton, M.; Baldisseri, D. M.; Hadad, C. M.; Peczu, M. W. *J. Org. Chem.* **2005**, *70*, 24–38.